Freshwater Ciliates (Protozoa, Ciliophora) from Argentina: An Annotated and Updated Compilation

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INTRODUCTION

The phylum Ciliophora is considered to be among the most diverse from kingdom Protozoa [1] and includes heterotroph, autotroph, and mixotroph microorganisms. Many species are euryoecious and have great adaptability against changing environmental conditions, being able to inhabit a wide range of different kinds of habitats, e.g., the commonly found planktonic oligotrich *Halteria grandinella* [2]. Among many morphological and physiological adaptations, the formation of resting cysts is a widespread phenomenon in ciliates, which enables them to survive during unfavorable environmental conditions, thus facilitating their dispersion to distant geographic regions. Nevertheless, the cysts have different properties depending on the region and habitat where they were formed [3].

The study of freshwater ciliates from Argentina began with the investigations of de la Rua [4], who studied the protozoa from pools in Capital Federal (Federal district), Buenos Aires. Seckt [5] and Martínez-Bustos [6] performed hydrobiological studies in Córdoba province, and enumerated the ciliates among other organisms from different kinds of aquatic ecosystems, such as ponds, streams and rivers. Carbonell [7] listed the ciliates found in the plankton, periphyton and mud samples from Río de la Plata, possibly from fluvial as well as brackish and marine zones. Later Balech [8] described a new ciliate genus and species from Capital Federal and Santa Fe province. After almost 20 years, devoided of information about these microorganisms, some researchers began to publish mainly new records of ciliates for Argentina [9-24], a new folliculinid genus and species [25], a new tintinnid species [26], and a new peritrich species [17]. Most of these investigations were based on live observations and supravital staining, without permanent slides collections available. Silver staining with Protargol and scanning electron microscopy was

employed for the first time in the country by Pettigrosso [27] and Barría de Cao [28], who surveyed planktonic ciliates from Bahía Blanca estuary in the Buenos Aires province. Concerning freshwater environments, Küppers [29] and Küppers et al. [30–34] described the ciliates from a temporary pond near the locality of Poblet, in Buenos Aires province, by means of live observations and Protargol staining.

Ecological studies on freshwater ciliates are almost circumscribed to deep oligotrophic Andean lakes from Patagonia, although some investigations were carried out in Córdoba and Buenos Aires provinces as well [35-47]. In 1995, Foggetta revised the Argentinean ciliates from continental freshwaters and at that time, there were around 100 cited species, with most of the genera being monospecific. Later, Küppers [29] found 70 freshwater species in only one temporary pond during a biannual investigation. The aim of the present contribution is to update the compilation of Argentinean freshwater ciliates, with notes on their habitat and distribution.

MATERIALS AND METHODS

The following species list was produced by examining the literature that is specifically referred to ciliates, and also faunistic or ecological studies where ciliates were mentioned for freshwater environments from Argentina. Carbonell [7] cited many freshwater as well as marine species for the Río de la Plata, which was included in the present contribution, since the author did not mention whether the samplings were conducted in fluvial or brackish-marine zones. On the contrary, the species cited by Souto [26] and Kogan [48] for Río de la Plata estuary were excluded since the samplings were conducted in brackish or marine zones. Some species mentioned by Claps et al. [49] for the Salado River basin are also commonly found in marine habitats, since this river has high concentration of dissolved salts due to the basin nature. Küppers [29] recorded the ciliates that developed in the laboratory from resting cysts in the dry sediments of the bed from a temporary pond that dries mainly during the summer. Some of these species were included only when cited by other authors in further freshwater environments.

The taxonomy is according to Lynn [50] and specific taxonomic papers. Abbreviations are as follows: BA, Buenos Aires province; CH, Chubut province; CO, Córdoba province; NE, Neuquén province; RN, Río Negro province; SF, Santa Fe province.

RESULTS

The following 209 species listed below were cited for freshwater environments in Argentina. The highest species richness occurs in the orders Sessilida (26% of the total number of species), Heterotrichida (9%), Haptorida (9%), and Sporadotrichida (7%). In Buenos Aires province, 193 species (92%) were recorded, while 22% of the total number of species were found in Córdoba province (Fig. 4.1), with many of them (41 species) being present in both provinces. Less than 4% of the species are distributed in the remaining vast territory of Argentina.

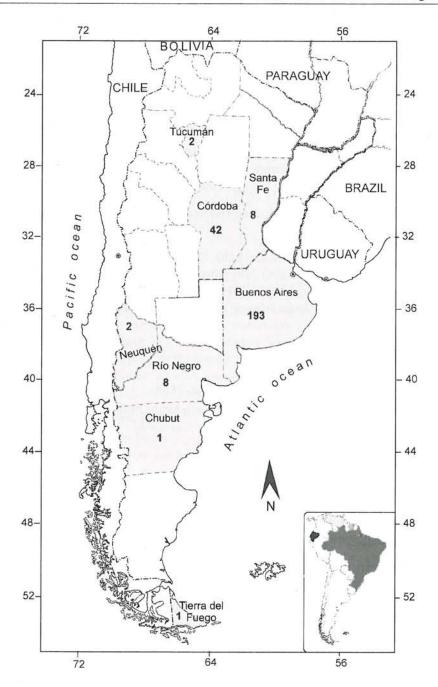


Fig. 4.1 Records of ciliates in Argentina. Surveyed provinces are highlighted in grey and numbers correspond to the total species recorded in each province.

Phylum Ciliophora Doflein, 1901

Class Heterotrichea Stein, 1859

Order Heterotrichida Stein, 1859

Family BLEPHARISMIDAE Jankowski in Small & Lynn, 1985

Blepharisma Perty, 1849

Blepharisma americanum (Suzuki, 1954) Hirshfield, Isquith & Bhandary, 1965

Recorded near Poblet (BA), in plankton and rewetted soil samples from a temporary pond [29, 34]. Observation techniques: *in vivo*, Protargol staining [51].

B. lateritium (Ehrenberg, 1831) Stein, 1859

Recorded in pools from Capital Federal, cited also as B. ovata [4]. Observation techniques: in vivo.

B. undulans Stein, 1867

Recorded in the periphyton from Las Mulas Lake, Chascomús (BA) [10]; in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); near Poblet (BA), in the plankton and rewetted soil samples from a temporary pond [29]. Observation techniques: *in vivo*, methyl green, vital staining, Chatton-Lwoff silver staining [52], ferric hematoxylin, Protargol staining [51], osmic acid vapours.

Family CLIMACOSTOMIDAE Repak, 1972

Climacostomum Stein, 1859

Climacostomum patulum Claparède & Lachmann, 1859

Recorded in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist). This species was found in marine environments by other authors [53]. Observation techniques: not indicated.

C. virens (Ehrenberg, 1838) Stein, 1859

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist). Observation techniques: *in vivo*.

Family CONDYLOSTOMATIDAE Kahl in Doflein & Reichenow, 1929

Linostomella Aescht in Foissner, Berger, & Schaumberg, 1999

Linostomella vorticella (Ehrenberg, 1833) Aescht in Foissner, Berger & Schaumburg, 1999

Recorded in pools from Capital Federal, cited as *Condylostoma vorticella* [4]; near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) and rewetted soil samples from a temporary pond [29] (checklist). Observation techniques: *in vivo*, Protargol staining [51].

Family FOLLICULINIDAE Dons, 1914

Ascobius Henneguy, 1884

Ascobius lentus Henneguy, 1884

Recorded in Chascomús (BA), in the plankton from El Burro Lake [9] (empty shells); in the periphyton (*Azolla filiculoides*, *Salvinia rotundifolia*, *Ceratophyllum demersum* var. *oxycanthum*) from Yalca, El Burro, and Chis Chis Lakes (BA) [11]. Observation techniques: *in vivo*.

Botticula Dioni, 1972

Botticula ringueleti Dioni, 1972

Recorded in Santa Fe province, in the periphyton from Salado River in Santo Tomé (*Pistia* sp., *Eichhornia* sp.), from "madrejón" Don Felipe in Colastiné Sur River, and from 'madrejón' El Alemán in El Vado island (*Pistia* sp., *Eichhornia* sp., *Salvinia* sp.) [25]. Observation techniques: *in vivo*.

Folliculina Lamarck, 1816

Folliculina boltoni Kent, 1881

Recorded in De Los Sapos island (SF), in the periphyton (*Pistia* sp.), cited as *Diafolliculina thomseni* [25]; in the plankton from the Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, formol-fixed.

Family SPIROSTOMIDAE Stein, 1867

Spirostomum Ehrenberg, 1834

Spirostomum ambiguum (O. F. Müller, 1786) Ehrenberg, 1835

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers [5] (checklist); in the plankton, periphyton, and mud samples from Río de la Plata [7] (checklist); in Chascomús (BA), in the periphyton from Las Mulas Lake [10]; in the plankton from San Miguel del Monte Lake (BA) [54] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin.

S. teres Claparède & Lachmann, 1858

Recorded in pools from Capital Federal [4]; in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [22, 46] (checklist). Observation techniques: *in vivo*, formol-fixed.

Family STENTORIDAE Carus, 1863

Stentor Oken, 1815

Stentor araucanus Foissner & Wölfl, 1994

Recorded in Río Negro and Chubut provinces, in the plankton from ultraoligotrophic lakes: Nahuel Huapi, Gutiérrez, Moreno (RN), and Vintter (CH) possibly coexisting with *S. amethystinus* [55]; in Neuquén and Río Negro provinces, in the plankton from oligotrophic lakes: Correntoso (NE), Nahuel Huapi, Moreno Este, Moreno Oeste, Gutiérrez, Mascardi, and Guillelmo (RN) [36, 40, 43-45]. Observation techniques: *in vivo*, silver carbonate staining [56].

S. barretti Barret, 1870

Recorded in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist). Observation techniques: not indicated.

S. coeruleus (Pallas, 1766) Ehrenberg, 1831

Recorded in pools from Capital Federal [4]; in the plankton and periphyton from ponds, streams, and rivers [5] (checklist); in artificial pools from La Plata (BA) [58]; in the benthos from Río Primero (CO) [6]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata (BA) [7] (checklist); in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); in the plankton from Luján river (BA) [59] (checklist); near Poblet (BA), in the plankton from a temporary pond [29] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, vital staining, formol-fixed, Protargol staining [51].

S. igneus Ehrenberg, 1838

Recorded near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29]. Observation techniques: *in vivo*, methyl-green-pyronin, Protargol staining [51].

S. multiformis (O. F. Müller, 1786) Ehrenberg, 1838

Recorded near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist). Observation techniques: *in vivo*, methyl-green-pyronin, Protargol staining [51].

S. niger (O. F. Müller, 1773) Ehrenberg, 1831

Recorded in Córdoba province, in the plankton and periphyton from ponds and streams (not specifically indicated) [5] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: *in vivo*, formol-fixed.

S. polymorphus (O. F. Müller, 1773) Ehrenberg, 1830

Recorded in pools from Capital Federal [4]; in Buenos Aires and Córdoba provinces, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton and benthos from Río Primero (CO) [6]; in periphyton and mud samples from Río de la Plata [7] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

S. roeselii Ehrenberg, 1835

Recorded in the plankton from Luján and Arrecifes rivers and Tala stream (BA) [59] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [22, 46] (checklist); near Poblet (BA), in the plankton from a temporary pond [29]; in the plankton from San Miguel del Monte Lake (BA) [54] (checklist); in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, formol-fixed, methyl-green-pyronin, Protargol staining [51].

Class SPIROTRICHEA Bütschli, 1889 Order Euplotida Small & Lynn, 1985 Family ASPIDISCIDAE Ehrenberg, 1830

Aspidisca Ehrenberg, 1830

Aspidisca cicada (O. F. Müller, 1786) Claparède & Lachmann, 1858

Recorded in pools from Capital Federal [4]; in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18]; in the plankton from Río de la Plata estuary [37] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); near Poblet (BA), in the plankton and periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist). Cited as *A. costata* by de la Rua [4] and Zaleski & Claps [46]. Observation techniques: *in vivo*, vital staining, methyl-green, neutral red, silver staining [60], formol-fixed, lugol fixed.

A. lynceus (O. F. Müller, 1773) Ehrenberg, 1830

Recorded in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18]; in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: *in vivo*, vital staining, formol-fixed.

A. turrita (Ehrenberg, 1838) Claparède & Lachmann, 1858

Recorded in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18]. Observation techniques: in vivo, vital staining.

Family EUPLOTIDAE Ehrenberg, 1838

Euplotes Ehrenberg in Hemprich & Ehrenberg, 1831

Euplotes charon (O. F. Müller, 1773) Ehrenberg, 1830

Recorded in pools from Capital Federal [4]. Observation techniques: in vivo.

E. moebiusi Kahl, 1932

Recorded in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist). Observation techniques: in vivo, vital staining.

Euplotoides Borror & Hill, 1995

Euplotoides eurystomus (Wrzesniowski, 1870) Borror & Hill, 1995

Recorded in Chascomús, in the periphyton from Vitel Lake (BA) (Lemna minima), and from Espín stream (Pistia stratiotes) (SF) [10]; in Punta Atalaya, Magdalena (BA), in the periphyton (Scirpus californicus) from Río de la Plata estuary [61] (checklist); in La Plata (BA), in the plankton from Rodríguez stream [62] (checklist); in the plankton from Luján River (BA) [59] (checklist); near Poblet (BA), in the plankton and periphyton (Alternanthera philoxeroides, Ludwigia peploides) from a temporary pond [29] (checklist). Cited as Euplotes eurystomus in most cases, or Euplotes plumipes [10]. Observation techniques: in vivo, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin, formol-fixed, Protargol staining [51], osmic acid vapours.

E. patella (O. F. Müller, 1773) Borror & Hill, 1995

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in Chascomús (BA), in the periphyton (*Lemna* sp.) from Vitel Lake [10]; in La Plata (BA), in the plankton from Rodríguez stream [62] (checklist); in the plankton from Luján River and Pescado stream (BA) [59] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); near Poblet (BA), in the plankton and periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29]

(checklist); in the plankton from Salado River basin (BA) [49] (checklist). Cited as *Euplotes patella* in most cases. Observation techniques: *in vivo*, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin, formol-fixed, Protargol staining [51], osmic acid vapours.

Euplotopsis Borror & Hill, 1995

Euplotopsis affinis (Dujardin, 1841) Borror & Hill, 1995

Recorded in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [22, 46] (checklist). Observation techniques: *in vivo*, formol-fixed.

Family URONYCHIIDAE Jankowski, 1975

Diophrys Dujardin, 1841

Diophrys scutum (Dujardin 1841) Kahl, 1932

Recorded in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata, cited as *D. grandis* [7] (checklist). This species was cited for marine environments by other authors [63]. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

Order Tintinnida Kofoid & Campbell, 1929

Family CODONELLIDAE Kent, 1881

Codonella Haeckel, 1873

Codonella cratera (Leidy, 1887) Imhof, 1885

Recorded in Capital Federal, in the periphyton (*Vaucheria* sp.) from ponds, streams, and rivers (not specifically indicated), cited as *C. lacustris* [5] (checklist); in the plankton from Río de la Plata estuary, the Paraná River delta, and artificial lakes from Capital Federal [12]; in the plankton from Baradero, Arrecifes, Ramallo, and Arroyo del Medio rivers (BA) [59] (checklist); in the plankton from Río de la Plata estuary [37] (checklist); in the plankton from San Miguel del Monte Lake (BA) [54]; in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, methyl-green, neutral red, formol-fixed, silver staining [60].

Tintinnopsis Stein, 1867

Tintinnopsis cylindrata Kofoid & Campbell, 1929

Recorded in the plankton from Río de la Plata estuary [37] (checklist). Observation techniques: *in vivo*, methyl-green, neutral red, silver staining [60].

T. fimbriata Meunier, 1919

Recorded in the plankton from Salado River basin (BA), cited as *Codonaria fimbriata* [49] (checklist). Observation techniques: *in vivo*, formol-fixed.

T. rioplatensis Souto, 1973

Recorded in the plankton from internal and intermediate fluvial zones of Río de la Plata estuary, Paraná River delta, and artificial lakes from Capital Federal [12]; in the plankton from Río de la Plata estuary [37] (checklist). Observation techniques: *in vivo*, methyl-green, neutral red, formol fixed, silver staining [60].

Family TINTINNIDIIDAE Kofoid & Campbell, 1929

Tintinnidium Kent, 1881

Tintinnidium fluviatile (Stein, 1863) Kent, 1881

Recorded in Córdoba province, in the plankton and periphyton from ponds and streams (not specifically indicated) [5] (checklist); in the plankton from Río de la Plata estuary and artificial lakes in Capital Federal [12]; in Punta Atalaya, Magdalena (BA), in the periphyton (*Scirpus californicus*) from Río de la Plata [61] (checklist); in the plankton from Baradero, Arrecifes, and Arroyo del Medio rivers, and Giles stream (BA) [59] (checklist); near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist); in the plankton from San Miguel del Monte Lake (BA) [54] (checklist); in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, formol-fixed, Lugol-fixed.

T. pusillum Entz, 1909

Recorded in the plankton from the fluvial zone of Río de la Plata estuary and in artificial lakes from Capital Federal [12]. Observation techniques: *in vivo*, formol-fixed.

Order Choreotrichida Small & Lynn, 1985

Family STROBILIDIIDAE Kahl in Doflein & Reichenow, 1929

Pelagostrobilidium Petz, Song, & Wilbert, 1995

Pelagostrobilidium wilberti Küppers, Lopretto & Claps, 2006

Recorded in Magdalena and Punta Lara (BA), in the plankton from temporary ponds [29, 30]. Observation techniques: *in vivo*, Protargol staining [51].

Rimostrombidium Jankowski, 1978

Rimostrombidium brachykinetum Krainer, 1995

Recorded near Poblet (BA), in the plankton from a temporary pond [29] (checklist). Observation techniques: in vivo.

R. humile (Penard, 1922) Petz & Foissner, 1992

Recorded in the plankton from the oligotrophic Morenito Lake (RN), cited as *Strobilidium humile* [24]. Observation techniques: *in vivo*, Lugol-fixed.

R. lacustris (Foissner, Skogstad & Pratt, 1988) Petz & Foissner, 1992

Recorded in the plankton from the oligotrophic Morenito Lake (RN), cited as *Strobilidium lacustris* [24]. Observation techniques: *in vivo*, Lugol-fixed.

Strobilidium Schewiakoff, 1893

Strobilidium caudatum (Fromentel, 1876) Foissner, 1987

Recorded in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA), cited as *S. gyrans* [18] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA), cited as *Strombilidium gyrans* [46] (checklist); near Poblet (BA), in the plankton and periphyton (*Althernanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29, 31]. Observation techniques: *in vivo*, vital staining, methyl-green-pyronin, formol-fixed.

Order Stichotrichida Fauré-Fremiet, 1961

Family SPIROFILIDAE von Gelei, 1929

Hypotrichidium Ilowaisky, 1921

Hypotrichidium conicum Ilowaisky, 1921

Recorded near Poblet (BA), in the plankton and rewetted soil samples from a temporary pond [29, 31]. Observation techniques: in vivo, Protargol staining [51].

Stichotricha Perty, 1849

Stichotricha secunda Perty, 1849

Recorded in pools from Capital Federal [4]; near Poblet (BA), in the plankton and periphyton (Alternanthera philoxeroides, Ludwigia peploides) from a temporary pond [29] (checklist). Observation techniques: in vivo, Protargol staining [51], Lugol-fixed.

Order Sporadotrichida Fauré-Fremiet, 1961

Family HALTERIIDAE Claparède & Lachmann, 1858

Halteria Dujardin, 1841

Halteria grandinella (O. F. Müller, 1773) Dujardin, 1841

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in Chascomús (BA), in the periphyton (Lemna minima) from Vitel Lake [10]; in Punta Atalaya, Magdalena (BA), in the periphyton (Scirpus californicus) from Río de la Plata [61] (checklist); in La Plata (BA), in the plankton from Rodríguez stream [62] (checklist); in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); in the plankton from Río de la Plata estuary [37] (checklist); near Poblet (BA), in the plankton, periphyton (Alternanthera philoxeroides, Ludwigia peploides), and rewetted soil samples from a temporary pond [29, 31]; in the plankton from San Miguel del Monte Lake (BA) [54]; in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: in vivo, Bouin-fixed, osmic acid vapours, methyl-green, neutral red, silver staining [60], Chatton-Lwoff silver staining [52], ferric hematoxylin, formol-fixed, Protargol staining [51].

Family OXYTRICHIDAE Ehrenberg, 1830

Apoamphisiella Foissner, 1997

Apoamphisiella tihanyiensis (Gellért & Tamás, 1958) Foissner, 1997

Recorded near Poblet (BA), in the plankton from a temporary pond [29]. Probably misidentified due to the particular pattern of dorsal rows of bristles. Observation techniques: in vivo, Protargol staining [51].

Cyrtohymena Foissner, 1989

Cyrtohymena candens (Kahl, 1932) Foissner, 1989

Recorded near Poblet (BA), in the plankton from a temporary pond [29]. Observation techniques: in vivo, Protargol staining [51].

Histriculus Corliss, 1960

Histriculus histrio (O. F. Müller, 1773) Corliss, 1960

Recorded in Capital Federal and in Córdoba province in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated), cited as *Stylonychia histrio* [5] (checklist); near Poblet (BA), in the plankton from a temporary pond [29]. Observation techniques: *in vivo*, Protargol staining [51].

Oxytricha Bory de St. Vincent in Lamouroux, Bory de St. Vincent & Deslongchamps, 1824 Oxytricha fallax Stein, 1859

Recorded in Chascomús (BA), in the periphyton (*Azolla filiculoides*) from Yalca Lake [10]; in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist). Observation techniques: *in vivo*, methyl-green, vital staining, Chatton-Lwoff silver staining [52], ferric hematoxylin, osmic acid vapours.

Paraurostyla Borror, 1972

Paraurostyla weissei (Stein, 1859) Borror, 1972

Recorded in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated), cited as *Urostyla weissei* [5] (checklist). Observation techniques: not indicated.

Pleurotricha Stein, 1859

Pleurotricha lanceolata (Ehrenberg, 1835) Stein, 1859

Recorded near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Typha* sp.) from a temporary pond [29]. Observation techniques: *in vivo*, Protargol staining [51].

Rubrioxytricha Berger, 1999

Rubrioxytricha ferruginea (Stein, 1859) Berger, 1999

Recorded in pools from Capital Federal [4]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). Cited as *Oxytricha ferruginea*. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

Stylonychia Ehrenberg, 1830

Stylonychia lemnae Ammermann & Schlegel, 1983

Recorded near Poblet (BA), in the plankton, periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*), and rewetted soil samples from a temporary pond [29, 34]. Observation techniques: *in vivo*, Protargol staining [51].

S. mytilus (O. F. Müller, 1773) Ehrenberg, 1830

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in Punta Atalaya, Magdalena (BA), in the periphyton (*Scirpus californicus*) from Río de la Plata [60] (checklist); in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); near Poblet (BA), in plankton and rewetted soil samples from a temporary pond [29]. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, formol-fixed, vital staining, Protargol staining [51].

S. pustulata (O. F. Müller, 1786) Ehrenberg, 1835

Recorded in pools from Capital Federal [4]; in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the benthos from Río Primero (CO) [6]; in Berisso (BA), in the periphyton from a Palo Blanco pond [10]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). Observation techniques: in vivo, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin, Bouin-fixed, osmic acid vapours.

Tachysoma Stokes, 1887

Tachysoma chilensis (Bürger, 1905) Berger, 1999

Recorded in pools from Capital Federal [4]; in plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). Cited as Oxytricha chilensis. Observation techniques: in vivo, Bouin-fixed, osmic acid vapours.

T. pellionellum (O. F. Müller, 1773) Borror, 1972

Recorded in pools from Capital Federal [4]; in Córdoba province, in the plankton and periphyton from ponds and streams (not specifically indicated) [5] (checklist) and in the plankton and benthos from Río Primero (CO) [6]; in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist). Cited as Oxytricha pellionella in most cases. Observation techniques: in vivo, vital staining.

Urosoma Kowalewski, 1882

Urosoma caudata (Ehrenberg, 1833) Berger, 1999

Recorded in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist). Observation techniques: in vivo, vital staining.

Order Urostylida Jankowski, 1979

Family UROSTYLIDAE Bütschli, 1889

Diaxonella Jankowski, 1979

Diaxonella pseudorubra (Kaltenbach, 1960) Berger, 2006

Recorded near Poblet (BA), in periphyton (Alternanthera philoxeroides, Ludwigia peploides) and rewetted soil samples from a temporary pond, cited as D. trimarginata [29] (checklist). Observation techniques: in vivo, Protargol staining [51].

Uroleptus Ehrenberg, 1831

Uroleptus musculus (Kahl, 1932) Foissner, Blatterer, Berger & Kohmann, 1991

Recorded in pools from Capital Federal [4]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); near Poblet (BA), in rewetted soil samples from a temporary pond [29]. Observation techniques: in vivo, Bouin-fixed, osmic acid vapours, Protargol staining [51].

U. piscis (O. F. Müller, 1773) Ehrenberg, 1831

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton and benthos from Río Primero (CO) [6]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); near Poblet (BA), in the plankton from a temporary pond, cited as *U. limnetis* [29] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, Protargol staining [51].

Urostyla Ehrenberg, 1830

Urostyla grandis Ehrenberg, 1830

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

Order Strombidiida Petz & Foissner, 1992

Family STROMBIDIIDAE Fauré-Fremiet, 1970

Limnostrombidium Krainer, 1995

Limnostrombidium pelagicum (Kahl, 1932) Krainer, 1995

Recorded near Poblet (BA), in the plankton and periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29, 31]. Observation techniques: *in vivo*, Protargol staining [51], Lugol-fixed.

L. viride (Stein, 1867) Krainer, 1995

Recorded in Córdoba province, in the plankton and periphyton from ponds and streams (not specifically indicated) [5] (checklist); in the plankton from the oligotrophic Morenito Lake (RN) [24]; near Poblet (BA), in the plankton and periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29, 31]. Cited as *Strombidium viride* in most cases. Observation techniques: *in vivo*, Protargol staining [51], Lugol-fixed.

Pelagostrombidium Krainer, 1991

Pelagostrombidium mirabile (Penard, 1916) Krainer, 1991

Recorded near Poblet (BA), in the plankton and periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29, 31]. Observation techniques: *in vivo*, Protargol staining [51], Lugol-fixed.

Strombidium Claparède & Lachmann, 1859

Strombidium elegans Florentin, 1901

Recorded in pools from Capital Federal, cited as *Strombidium armatum* [4]. This species was found in marine environments by other authors [63]. Observation techniques: *in vivo*.

Class ARMOPHOREA Lynn, 2004

Order Armophorida Jankowski, 1964

Family CAENOMORPHIDAE Poche, 1913

Caenomorpha Perty, 1852

Caenomorpha medusula Perty, 1852

Recorded in the plankton from Cassaffousth reservoir (CO) [21]. Observation techniques: in vivo, methyl-green, formol-fixed.

Family METOPIDAE Kahl, 1927

Brachonella Jankowski, 1964

Brachonella spiralis (Smith, 1897) Jankowski, 1964

Recorded in the plankton from Cassaffousth reservoir (CO) [21]; near Poblet (BA), in the plankton from a temporary pond [29] (checklist). Observation techniques: in vivo, methyl-green, formolfixed, Lugol-fixed.

Metopus Claparède & Lachmann, 1858

Metopus es (O. F. Müller, 1776) Lauterborn, 1916

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton from Cassaffousth reservoir (CO) [21]. Cited as M. sigmoides in most cases. Observation techniques: in vivo, methyl-green, formol-fixed.

Class LITOSTOMATEA Small & Lynn, 1981

Order Haptorida Corliss, 1974

Family DIDINIIDAE Poche, 1913

Didinium Stein, 1859

Didinium nasutum (O. F. Müller, 1773) Stein, 1859

Recorded in pools from Capital Federal [4]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in Berisso (BA), in the periphyton from a Palo Blanco pond [10]; in Punta Atalaya, Magdalena (BA), in the periphyton (Scirpus californicus) from Río de la Plata [60] (checklist); in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the plankton from Luján River and Pescado stream (BA) [58] (checklist); in the plankton from Río de la Plata estuary [37]; near Poblet (BA), in the periphyton (Alternanthera philoxeroides, Ludwigia peploides) from a temporary pond [29]; in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: in vivo, Bouin-fixed, osmic acid vapours, methyl-green, neutral red, Chatton-Lwoff silver staining [52], ferric hematoxylin, silver staining [59], Protargol staining [51], formol-fixed.

Monodinium Fabre-Domergue, 1888

Monodinium cf. balbiani

Recorded near Poblet (BA), in the periphyton (Alternanthera philoxeroides, Ludwigia peploides) and rewetted soil samples from a temporary pond [29] (checklist). Observation techniques: in vivo, Protargol staining [51].

Family LACRYMARIIDAE de Fromentel, 1876

Lacrymaria Bory de St. Vincent, 1824

Lacrymaria olor (O. F. Müller, 1786) Bory de Saint-Vincent, 1824

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the

plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in Chascomús (BA), in the periphyton (Salvinia rotundifolia) from Yalca Lake [10]; in Punta Atalaya, Magdalena (BA), in the periphyton (Scirpus californicus) from Río de la Plata [60] (checklist); in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist); in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); near Poblet (BA), in the plankton and periphyton (Alternanthera philoxeroides, Ludwigia peploides) from a temporary pond [29] (checklist). Observation techniques: in vivo, Bouin-fixed, osmic acid vapours, methyl-green, vital staining, Chatton-Lwoff silver staining [52], ferric hematoxylin, Protargol staining [51], Lugol-fixed, formol-fixed.

Phialina Bory de St. Vincent, 1824

Phialina coronata (Claparède & Lachmann, 1859) Foissner, 1987

Recorded in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata, cited as Lacrymaria coronata [7] (checklist). This species is usually found in marine environments [64]. Observation techniques: in vivo, Bouin-fixed, osmic acid vapours.

P. pupula (O. F. Müller, 1786) Foissner, 1983

Recorded in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata, cited as Lacrymaria elliptica [7] (checklist); in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA), cited as L. pupula [46] (checklist). This species was also cited for marine environments by other authors [53]. Observation techniques: in vivo, Bouin-fixed, osmic acid vapours, formol-fixed.

P. salinarum (Kahl, 1928)

Recorded in Buenos Aires province, in the plankton from Salado River basin, cited as Lacrymaria salinarum [49] (checklist). Observation techniques: in vivo; formol-fixed.

P. vermicularis (O. F. Müller-Ehrenberg, 1831) Foissner, 1983

Recorded in pools from Capital Federal (4); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata, cited as Lacrymaria metabolica [7] (checklist). This species was cited for marine environments by other authors [53]. Observation techniques: in vivo, Bouin-fixed, osmic acid vapours.

Family SPATHIDIIDAE Kahl in Doflein & Reichenow, 1929

Spathidium Dujardin, 1841

Spathidium spathula (O. F. Müller, 1773) Dujardin, 1841

Recorded in Capital Federal, in pools [4]. Observation techniques: in vivo.

Epispathidium Foissner, 1984

Epispathidium cf. amphoriforme

Recorded near Poblet (BA), in the plankton from a temporary pond [29] (checklist). Observation techniques: in vivo, Protargol staining [51].

Teuthophrys Chatton & de Beauchamp, 1923

Teuthophrys trisulca africana Dragesco & Dragesco-Kernéis, 1986

Recorded near Poblet (BA), in the plankton and rewetted soil samples from a temporary pond [29, 30]. Observation techniques: *in vivo*, Protargol staining [51].

Family TRACHELIIDAE Ehrenberg, 1838

Dileptus Dujardin, 1841

Dileptus anser (O. F. Müller, 1786) Dujardin, 1841

Recorded in pools from Capital Federal [4]; in Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist) and benthos from Río Primero [6]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist). Cited as *Lionotus anser* in most cases. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, vital staining.

Monilicaryon Jankowski, 1967

Monilikaryon monilatus (Stokes, 1886) Jankowski, 1967

Recorded near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist). Observation techniques: *in vivo*; Protargol staining [51].

Paradileptus Wenrich, 1929

Paradileptus elephantinus (Švec, 1897) Kahl, 1931

Recorded in the plankton from the oligotrophic Morenito Lake (RN) [24] and El Trébol Lake (RN) [38]; near Poblet (BA), in the plankton from a temporary pond [29] (checklist). Observation techniques: *in vivo*, Lugol-fixed, Protargol staining [51].

Trachelius Schrank, 1803

Trachelius ovum (Ehrenberg, 1831) Ehrenberg, 1838

Recorded near Poblet (BA), in the plankton from a temporary pond [29] (checklist). Observation techniques: *in vivo*, Protargol staining [51].

Family TRACHELOPHYLLIDAE Kent, 1882

Acaryophrya André, 1915

Acaryophrya cf. mamillata

Recorded near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist). Observation techniques: Protargol staining [51].

Enchelyodon Claparède & Lachmann, 1859

Enchelyodon laevis (Quennerstedt, 1867) Dragesco, 1960

Recorded in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata, cited as *Lagynus laevis* [7] (checklist). This species was found by other authors in marine environments [53]. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

Epitholiolus Foissner, Agatha, & Berger, 2002

Epitholiolus chilensis (Bürger, 1906) Foissner, Agatha & Berger, 2002

Recorded in pools from Capital Federal [4]; in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). Cited as Lacrymaria chilensis. Observation techniques: in vivo, Bouin-fixed, osmic acid vapours.

Lagynophrya Kahl, 1927

Lagynophrya cf. rostrata

Recorded near Poblet (BA), in the periphyton (Alternanthera philoxeroides, Ludwigia peploides) from a temporary pond [29] (checklist). Observation techniques: Protargol staining [51].

Order Pleurostomatida Schewiakoff, 1896

Family AMPHILEPTIDAE Bütschli, 1889

Amphileptus Ehrenberg, 1830

Amphileptus carchesii Stein, 1867

Recorded in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist). Observation techniques: not indicated.

A. claparedii Stein, 1867

Recorded in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist). Observation techniques: in vivo, vital staining.

A. pleurosigma (Stokes, 1884) Foissner, 1984

Recorded in the plankton from Río de la Plata estuary [37] (checklist); in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA), cited as Hemiophrys pleurosigma [46] (checklist). Observation techniques: in vivo, methyl-green, neutral red, silver staining [59], formol-fixed.

Family LITONOTIDAE Kent, 1882

Litonotus Wresniowski, 1870

Litonotus carinatus Stokes, 1885

Recorded in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist). Observation techniques: in vivo, vital staining.

L. fasciolla (Ehrenberg, 1830) Song & Wilbert, 1989

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated), cited as Loxophyllum fasciolla [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata, cited as Lionotus fasciolla [7] (checklist); in Chascomús (BA), in the periphyton (Salvinia rotundifolia) from Yalca Lake [10]; in Punta Atalaya, Magdalena (BA), in the periphyton (Scirpus californicus) from Río de la Plata [60] (checklist); in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: in vivo,

Bouin-fixed, osmic acid vapours, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin, formol-fixed.

L. varsaviensis Wrzesniowski, 1870

Recorded in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA), cited as Hemiophrys bivacuolata [46] (checklist). Observation techniques: in vivo, formol-fixed.

Loxophyllum Dujardin, 1841

Loxophyllum helus (Stokes, 1884) Penard, 1922

Recorded in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: in vivo, formol-fixed.

Order Cyclotrichiida Jankowski, 1980 Incertae sedis

Family MESODINIIDAE Jankowski, 1980

Askenasia Blochmann, 1895

Askenasia chlorelligera Krainer & Foissner, 1990

Recorded near Poblet (BA), in the periphyton (Alternanthera philoxeroides) from a temporary pond [29] (checklist). Observation techniques: Protargol staining [51].

A. volvox (Eichwald, 1852) Kahl, 1930

Recorded in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); near Poblet (BA), in the plankton and periphyton (Alternanthera philoxeroides, Ludwigia peploides) from a temporary pond [29, 31]. Observation techniques: in vivo, formol-fixed, Protargol staining [51], Lugol-fixed.

Rhabdoaskenasia Krainer & Foissner, 1990

Rhabdoaskenasia minima Krainer & Foissner, 1990

Recorded near Poblet (BA), in the plankton from a temporary pond [29, 31]. Observation techniques: in vivo, Protargol staining [51].

Class PHYLLOPHARYNGEA de Puytorac et al., 1974

Subclass Cyrtophoria Fauré-Fremiet in Corliss, 1956

Order Chlamydodontida Deroux, 1976

Family CHILODONELLIDAE Deroux, 1970

Chilodonella Strand, 1928

Chilodonella megalotrochae (Stokes, 1884)

Recorded in Berisso (BA), in the periphyton from Palo Blanco pond, cited as Chilodon megalotrochae [10], Observation techniques: in vivo, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin.

Ch. uncinata (Ehrenberg, 1838) Strand, 1928

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated), cited as Chilodon uncinatus [5] (checklist); in Punta Atalaya, Magdalena (BA), in the periphyton (Scirpus californicus) from Río de la Plata [60] (checklist); in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist); in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); near Poblet (BA), in plankton and rewetted soil samples from a temporary pond [29] (checklist); in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: in vivo, vital staining, formol-fixed, Protargol staining [51].

Pseudochilodonopsis Foissner, 1979

Pseudochilodonopsis piscatoris (Blochmann, 1895) Foissner, 1979

Recorded near Poblet (BA), in the periphyton (Alternanthera philoxeroides, Ludwigia peploides) from a temporary pond [29]. Observation techniques: in vivo, Protargol staining [51], Lugol-fixed.

Trithigmostoma Jankowski, 1967

Trithigmostoma cucullulus (O. F. Müller, 1786) Jankowski, 1967

Recorded in pools from Capital Federal [4]; in Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist); in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Cited as Chilodon cucullulus [4, 5, 7] or Chilodonella cucullus [18, 46]. Observation techniques: in vivo, Bouin-fixed, osmic acid vapours, formol fixed, vital staining.

Family GASTRONAUTIDAE Deroux, 1994

Gastronauta Engelmann in Bütschli, 1889

Gastronauta membranaceus Bütschli, 1889

Recorded near Poblet (BA), in the plankton and periphyton (Alternanthera philoxeroides, Ludwigia peploides) from a temporary pond [29] (checklist). Observation techniques: in vivo, Protargol staining [51], Lugol-fixed.

Subclass Suctoria Claparède & Lachmann, 1858

Order Exogenida Collin, 1912

Family METACINETIDAE Bütschli, 1889

Metacineta Bütschli, 1889

Metacineta cuspidata (Kellikott, 1885) Mathes, 1988

Recorded in the periphyton (Oedogonium sp., Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA), cited as Kellicotta cuspidata [22, 46] (checklist). Observation techniques: in vivo, formol-fixed.

Family PODOPHRYIDAE Haeckel, 1866

Podophrya Ehrenberg, 1834

Podophrya fixa (O. F. Müller, 1786) Ehrenberg, 1833

Recorded in pools from Capital Federal [4]; in Magdalena (BA), in the periphyton (Lemna sp.) from Las Víboras stream [16]; in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist); in the periphyton (Lemna sp.) from Luján River (BA) [16]; near Poblet (BA), in the plankton from a temporary pond [29] (checklist). Observation techniques: in vivo, vital staining, formol-fixed.

P. libera Perty, 1852

Recorded in pools from Capital Federal [4]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). Observation techniques: in vivo, Bouinfixed, osmic acid vapours.

Sphaerophrya Claparède & Lachmann, 1859

Sphaerophrya magna Maupas, 1881

Recorded in pools from Capital Federal [4]. Observation techniques: in vivo.

S. sol Metschnikoff, 1864

Recorded in pools from Capital Federal [4]. Observation techniques: in vivo.

Order Endogenida Collin, 1912

Family ACINETIDAE Stein, 1859

Acineta Ehrenberg, 1834

Acineta flava Kellicott, 1885

Recorded in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: in vivo, formol-fixed.

A. tuberosa (Pallas, 1766) Ehrenberg, 1833

Recorded in the periphyton from Luján River (BA) [15]; in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: in vivo, formol-fixed.

Acinetides Swarczewsky, 1928

Acinetides triangularis (Penard, 1920) Curds, 1985

Recorded in the periphyton (Oedogonium sp., Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [22, 46] (checklist). Observation techniques: in vivo, formol-fixed.

Suctorella Frenzel, 1891

Suctorella ciliata Frenzel, 1891

Recorded in muddy pools from Patagonia [65]. No further details available.

Family TOKOPHRYIDAE Jankowski in Small & Lynn, 1985

Tokophrya Bütschli, 1889

Tokophrya fasciculata (López-Ochoterena, 1964) Matthes, Guhl & Haider, 1988

Recorded in La Plata (BA), on Epistylis plicatilis from Rodríguez stream and in Magdalena (BA),

epibiont on *Drepanotrema* sp., (Gastropoda) from Las Víboras stream, cited as *Hypophrya fasciculata* [14, 61] (checklist); in the plankton and periphyton from Luján River (BA) [15, 58]. Observation techniques: *in vivo*, formol-fixed.

T. lemnarum (Stein, 1859) Entz, 1902

Recorded in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: in vivo, formol-fixed.

T. pyrum (Claparède & Lachmann, 1859) Butschli, 1889

Recorded in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: in vivo, formol-fixed.

T. quadripartita (Claparède & Lachmann, 1859) Bütschli, 1889

Recorded in pools from Capital Federal [4]; in the plankton from Nahuel Huapi Lake (RN) [66] (checklist); in the periphyton from Luján River (BA) [15]; in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, formol-fixed.

Order Evaginogenida Jankowski, 1978

Family DISCOPHRYIDAE Collin, 1912

Discophrya Lachmann, 1859

Discophrya elegans (Goodrich & Jahn, 1943) Matthes, Guhl & Haider, 1988

Recorded in the plankton and periphyton from Luján River (BA) [15, 58]. Cited as Multi-fasciculatum elegans. Observation techniques: in vivo, formol-fixed.

D. elongata (Claparede & Lachmann, 1858-1859) Collin 1911

Recorded in the plankton and periphyton from Luján River (BA) [15, 58]. Observation techniques: in vivo, formol-fixed.

Family HELIOPHRYIDAE Corliss, 1979

Heliophrya Saedeleer & Tellier, 1930

Heliophrya rotunda (Hentschel, 1916) Matthes, 1954

Recorded in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: *in vivo*, formol-fixed.

Class NASSOPHOREA Small & Lynn, 1981

Order Nassulida Jankowski, 1967

Family NASSULIDAE de Fromentel, 1874

Nassula Ehrenberg, 1834

Nassula ambigua Stein, 1854

Recorded in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

N. flava Claparede & Lachmann, 1859

Recorded in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la

Plata [7] (checklist). Observation techniques: in vivo, Bouin-fixed, osmic acid vapours.

Order Synhymeniida de Puytorac et al. in Deroux, 1978

Family ORTHODONELLIDAE Jankowski, 1968

Zosterodasys Deroux, 1978

Zosterodasys transversa (Kahl, 1928) Foisner, Berger & Kohmann, 1994

Recorded in the periphyton (*Pistia stratiotes*) from Espín stream (SF) [10]; in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist). Cited as *Chilodontopsis vorax*. Observation techniques: *in vivo*, methyl-green, vital staining, Chatton-Lwoff silver staining [52], ferric hematoxylin.

Class COLPODEA Small & Lynn, 1981

Order Bursariomorphida Fernández-Galiano, 1978

Family BURSARIIDAE Bory de St. Vincent, 1826

Bursaria O. F. Müller, 1773

Bursaria truncatella O. F. Müller, 1773

Recorded in La Plata, in the plankton from Rodríguez stream (BA) [61] (checklist); in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); near Poblet (BA), in rewetted soil samples from a temporary pond [29] (checklist). Observation techniques: *in vivo*, formol-fixed, vital staining, Protargol staining [51].

Order Colpodida de Puytorac et al., 1974

Family COLPODIDAE Bory de St. Vincent, 1826

Colpoda O.F. Müller, 1773

Colpoda cucullus (O. F. Müller, 1773) Gmelin, 1790

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); near Poblet (BA), in the plankton and rewetted soil samples from a temporary pond [29]. Observation techniques: *in vivo*, formol-fixed, Bouin-fixed, osmic acid vapours, Protargol staining [51].

C. inflata (Stokes, 1884) Kahl, 1931

Recorded near Poblet (BA), in the plankton from a temporary pond [29]. Observation techniques: *in vivo*, Protargol staining [51], Lugol-fixed.

C. steinii Maupas, 1883

Recorded in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); near Poblet (BA), in rewetted soil samples from a temporary pond [29]. Observation techniques: *in vivo*, Protargol staining [51].

Class PROSTOMATEA Schewiakoff, 1896

Order Prorodontida Corliss, 1974

Family BALANIONIDAE Small & Lynn, 1985

Balanion Wulff, 1919

Balanion planctonicum (Foissner, Oleksiv & Müller, 1990) Foissner, Berger & Kohmann, 1994

Recorded in the plankton from the oligotrophic Morenito Lake (RN) [24]. Observation techniques: in vivo, Lugol-fixed.

Family COLEPIDAE Ehrenberg, 1838

Coleps Nitzsch, 1827

Coleps bicuspis Noland, 1925

Recorded in the plankton from Río de la Plata estuary [37] (checklist). Observation techniques: *in vivo*, methyl-green, neutral red, silver staining [59].

C. hirtus (O. F. Müller, 1786) Nitzsch, 1827

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton and benthos from Río Primero (CO) [6]; in Chascomús (BA), in the periphyton (*Lemna minima*) from Vitel Lake [10]; in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); in the plankton from Río de la Plata estuary [37] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); near Poblet (BA), in the plankton and periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist); in the plankton from Lacombe Lake (BA) [67] (checklist); in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, methyl-green, neutral red, vital staining, Chatton-Lwoff silver staining [52], ferric hematoxylin, silver staining [59], formol-fixed, Lufol-fixed, Protargol staining [51], osmic acid vapours.

Family HOLOPHRYIDAE Perty, 1852

Holophrya Ehrenberg, 1831

Holophrya discolor Ehrenberg, 1833

Recorded in pools from Capital Federal, cited as P. rigidus [4]. Observation techniques: in vivo.

H. simplex Schewiakoff, 1893

Recorded in Chascomús (BA), in the periphyton from Vitel Lake [10]; in Punta Atalaya, Magdalena (BA), in the periphyton (*Scirpus californicus*) from Río de la Plata [60] (checklist); in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); in the plankton from San Miguel del Monte Lake (BA) [54]; in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, methyl-green, vital staining, Chatton-Lwoff silver staining [52], ferric hematoxylin, formol-fixed, osmic acid vapours.

H. teres (Ehrenberg, 1833) Foissner, Berger & Kohmann, 1994

Recorded in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Cited as *Prorodon teres*. Observation techniques: *in vivo*, formol-fixed.

Family PRORODONTIDAE Kent, 1881

Prorodon Ehrenberg, 1834

Prorodon edentatus Claparède & Lachmann, 1858

Recorded in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist). Observation techniques: not indicated.

P. taeniatus Blochmann, 1895

Recorded in the plankton from Río Tercero reservoir (CO) [35]. Observation techniques: *in vivo*, formol-fixed, scanning electron microscopy.

Family UROTRICHIDAE Small & Lynn, 1985

Bursellopsis Corliss, 1960

Bursellopsis nigricans (Lauterborn, 1894) Foissner, Berger & Schaumburg, 1999

Recorded in Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated), cited as *Holophrya nigricans* [5] (checklist). Observation techniques: not indicated.

Urotricha Claparède & Lachmann, 1859

Urotricha furcata Schewiakoff, 1892

Recorded in the plankton from the oligotrophic Morenito Lake (RN) [24]. Observation techniques: in vivo, Lugol-fixed.

Urotricha cf. globosa

Recorded near Poblet (BA), in the plankton from a temporary pond [29] (checklist). Observation techniques: *in vivo*, Lugol-fixed.

Class PLAGIOPYLEA Small & Lynn, 1985

Order Plagiopylida Jankowski, 1978

Family PLAGIOPYLIDAE Schewiakoff, 1896

Plagiopyla Stein, 1860

Plagiopyla nasuta Stein, 1860

Recorded in the plankton from Cassaffousth reservoir (CO) [21]. Observation techniques: in vivo, methyl-green, formol-fixed.

Incertae sedis in Class PLAGIOPYLEA

Order Odontostomatida Sawaya, 1940

Family EPALXELLIDAE Corliss, 1960

Saprodinium Lauterborn, 1908

Saprodinium dentatum (Lauterborn, 1901) Lauterborn, 1908

Recorded in the plankton from Cassaffousth reservoir (CO) [21]. Observation techniques: in vivo, methyl-green, formol-fixed.

Class OLIGOHYMENOPHOREA de Puytorac et al., 1974

Subclass Peniculia Fauré-Fremiet in Corliss, 1956

Order Peniculida Fauré-Fremiet in Corliss, 1956

Family FRONTONIIDAE Kahl, 1926

Disematostoma Lauterborn, 1894

Disematostoma buetschlii Lauterborn, 1894

Recorded near Poblet (BA), in the plankton from a temporary pond [29] (checklist). Observation techniques: in vivo.

Frontonia Ehrenberg, 1838

Frontonia atra (Ehrenberg, 1833) Bütschli, 1889

Recorded near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist). Observation techniques: *in vivo*, Lugol-fixed.

F. leucas (Ehrenberg, 1833) Ehrenberg, 1838

Recorded in pools from Capital Federal [4]; in Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton and benthos from Río Primero (CO) [6]; in the plankton, periphyton, and mud samples from Río de la Plata [7] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

Family NEOBURSARIDIIDAE Dragesco & Tuffrau, 1967

Neobursaridium Balech, 1941

Neobursaridium gigas Balech, 1941

Recorded in Capital Federal, in temporary ponds and in Santa Fe province, in Isla Candioti [8]. Observation techniques: *in vivo*, eosine-hematoxylin, silver nitrate.

Family PARAMECIIDAE Dujardin, 1840

Paramecium O. F. Müller, 1773

Paramecium aurelia O. F. Müller, 1773

Recorded in Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: *in vivo*, formol-fixed.

P. bursaria (Ehrenberg, 1831) Focke, 1836

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte

Lake (BA) [46] (checklist); near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist); in the plankton from San Miguel del Monte Lake (BA) [54] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, vital staining, formol-fixed.

P. caudatum Ehrenberg, 1833

Recorded in pools from Capital Federal [4]; in Capital Federal and Tandil Sierras (BA), and in Córdoba province in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton and benthos from Río Primero (CO) [6]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in Punta Atalaya, Magdalena (BA), in the periphyton (*Scirpus californicus*) from Río de la Plata [60] (checklist); in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [22, 46] (checklist); near Poblet (BA), in the plankton from a temporary pond [29] (checklist); in the plankton from San Miguel del Monte Lake (BA) [54] (checklist); in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, formol-fixed, Lugol-fixed.

Order Urocentrida Jankowski, 1980

Family UROCENTRIDAE Claparède & Lachmann, 1858

Urocentrum Nitzsch, 1827

Urocentrum turbo (O. F. Müller, 1786) Nitzsch, 1827

Recorded in pools from Capital Federal [4]; in Berisso (BA), in the periphyton from a Palo Blanco pond [10]; in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: *in vivo*, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin, formol-fixed, osmic acid vapours.

Subclass Scuticociliatia Small, 1967

Order Philasterida Small, 1967

Family CINETOCHILIDAE Perty 1852

Cinetochilum Perty, 1849

Cinetochilum margaritaceum (Ehrenberg, 1831) Perty, 1849

Recorded in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); in the plankton from Río de la Plata estuary [37] (checklist); near Poblet (BA), in the plankton and periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist). Observation techniques: *in vivo*, methyl-green, neutral red, silver staining [59], vital staining, Lugol-fixed, Protargol staining [51].

Family LOXOCEPHALIDAE Jankowski, 1964

Dexiotricha Stokes, 1885

Dexiotricha granulosa (Kent, 1881) Foissner, Berger & Kohmann, 1994

Recorded in pools from Capital Federal [4]; in Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist). Cited as *Loxocephalus granulosus*. Observation techniques: *in vivo*.

Family URONEMATIDAE Thompson, 1964

Uronema Dujardin, 1841

Uronema elegans Maupas, 1883

Recorded in Buenos Aires province, in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). This species was found in marine environment by other authors [53, 68]. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

U. marinum Dujardin, 1841

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton and benthos from Río Primero (CO) [6]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). This species was found in marine environments by other authors [69]. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

Order Pleuronematida Fauré-Fremiet in Corliss, 1956

Family CYCLIDIIDAE Ehrenberg, 1838

Cyclidium O.F. Müller, 1773

Cyclidium glaucoma O. F. Müller, 1773

Recorded in the periphyton (Azolla sp.) from Napostá Grande stream (BA), cited as Cyclidium cf. glaucoma [18] (checklist); near Poblet (BA), in the plankton and periphyton (Alternanthera philoxeroides, Ludwigia peploides) from a temporary pond [29] (checklist). Observation techniques: in vivo, vital staining, Lugol-fixed, Protargol staining [51].

Family PLEURONEMATIDAE Kent, 1881

Pleuronema Dujardin, 1841

Pleuronema crassum Dujardin, 1841

Recorded in pools from Capital Federal [4]; in Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in Berisso (BA), in the periphyton from a Palo Blanco pond [10]. Cited as *P. chrysalis* in most cases. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin.

Subclass Hymenostomatia Delage & Hérouard, 1896 Order Tetrahymenida Fauré-Fremiet in Corliss, 1956 Family GLAUCOMIDAE Corliss, 1971

Glaucoma Ehrenberg, 1830

Glaucoma scintillans Ehrenberg, 1830

Recorded in pools from Capital Federal [4]; in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); in Punta Lara (BA), in the periphyton (*Scirpus californicus*) from Río de la Plata [70] (checklist). Observation techniques: *in vivo*, vital staining, formol-fixed.

Family TETRAHYMENIDAE Corliss, 1952

Tetrahymena Furgason, 1940

Tetrahymena patula (0. F. Müller, 1786) Corliss, 1951

Recorded in Córdoba province, in the plankton and periphyton from ponds, streams, and rivers, cited as *Leucophrys patula* (not specifically indicated) [5] (checklist). Observation techniques: Not indicated.

T. pyriformis (Ehrenberg, 1830) Lwoff, 1947

Recorded in Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated), cited as *Glaucoma pyriformis* [5] (checklist); in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); in Tafi Viejo, Tucumán province, in a citric efluent from Salí River basin [71] (checklist); near Poblet (BA), in the plankton and periphyton (*Ludwigia peploides*) from a temporary pond [29] (checklist). Observation techniques: *in vivo*, vital staining, Lugol-fixed, neutral red, Protargol staining [51].

T. vorax (Kidder, Lilly & Claff, 1940) Kidder, 1941

Recorded in Tafi Viejo, Tucumán province, in a citric efluent from Salí River basin [71] (checklist). Observation techniques: *in vivo*, Lugol-fixed, neutral red.

Family TURANIELLIDAE Didier, 1971

Colpidium Stein, 1860

Colpidium colpoda (Losana, 1829) Stein, 1860

Recorded in pools from Capital Federal [4]; in Capital Federal and Sierras de Tandil (BA), and in Córdoba province in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in the periphyton (*Pistia stratiotes*) from Espín stream (SF) [10]; in the plankton from San Miguel del Monte Lake (BA) [54]. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin.

Subclass Peritrichia Stein, 1859 Order Sessilida Kahl, 1933 Family ASTYLOZOIDAE Kahl, 1935

Hastatella Erlanger, 1890

Hastatella radians Erlanger, 1890

Recorded in the plankton from Río de la Plata estuary [37] (checklist). Observation techniques: *in vivo*, methyl-green, neutral red, silver staining [59].

Family EPISTYLIDIDAE Kahl, 1933

Campanella Goldfuss, 1820

Campanella umbellaria (Linnaeus, 1758) Goldfuss, 1820

Recorded in the periphyton from a shallow pond in Berisso (BA) (*Lemna* sp.), and in the periphyton (*Ceratophyllum demersum*, *Scirpus californicus*, *Azolla filiculoides*, *Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [17, 46] (checklist); near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist). Observation techniques: *in vivo*, formol-fixed, methyl-green-pyronin.

Epistylis Ehrenberg, 1830

Epistylis aliciae Modenutti & Claps, 1986

Recorded in the periphyton (Scirpus californicus) from San Miguel del Monte Lake (BA) [17]. Observation techniques: in vivo.

E. articulata Fromentel, 1874

Recorded in the periphyton (*Myriophyllum quitense*) from San Miguel del Monte Lake (BA) [22]. Observation techniques: *in vivo*.

E. hentscheli Kahl, 1935

Recorded in the plankton (on debries) and periphyton (on *Enteromorpha* sp.) from San Miguel del Monte Lake (BA) [23]. Observation techniques: *in vivo*.

E. lacustris Imhoff, 1884

Recorded in Berisso (BA); in the periphyton from a Palo Blanco pond [10]. Observation techniques: *in vivo*, methyl-green, Chatton-Lwoff silver staining [52], ferric hematoxylin.

E. plicatilis Ehrenberg, 1831

Recorded in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in Chascomús (BA), in the periphyton from Las Mulas pond [10]; epibiont on *Pomacea canaliculata*, *P. insularum*, *P. scalaris* (Gastropoda) from the Middle Paraná River (SF) [72]; in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the plankton from Luján River (BA) [58] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); near Poblet (BA), in the plankton and periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29] (checklist); in Punta Lara (BA), in the periphyton (*Scirpus californicus*) from Río de la Plata [70] (checklist); in the plankton from San Miguel del Monte Lake (BA) [54]; in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, methyl-green, methyl-green-pyronin, Chatton-Lwoff silver staining [52], ferric hematoxylin, formol-fixed.

E. purneri Nenninger, 1948

Recorded in the periphyton (*Myriophyllum quitense*) from San Miguel del Monte Lake (BA) [22]. Observation techniques: *in vivo*.

E. cf. procumbens

Recorded near Poblet (BA), in the periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond, cited as *Epistylis* cf. *rotans* [29]. Observation techniques: *in vivo*, methylgreen-pyronin.

E. tubificis D'Udeckem 1864

Recorded in the periphyton (Myriophyllum quitense, Enteromorpha sp., Oedogonium sp.) from San Miguel del Monte Lake (BA) [23]. Observation techniques: in vivo.

E. cf. umbilicata

Recorded in Tierra del Fuego province, in the plankton from Cabecera Lake, near Fagnano Lake [73] (checklist). Observation techniques: fixed material.

E. vestita Stokes, 1887

Recorded in the periphyton (*Myriophyllum quitense*) from San Miguel del Monte Lake (BA) [23]. Observation techniques: *in vivo*.

Rhabdostyla Kent, 1881

Rhabdostyla pyriformis Perty, 1852

Recorded near Poblet (BA), epibiont on *Cypridopsis* sp. (Ostracoda) from a temporary pond [29] (checklist). Observation techniques: *in vivo*, methyl-green-pyronin.

Systylis hoffi Bresslau, 1919

Recorded in Punta Lara (BA), in the periphyton from Río de la Plata (Eichhornia crassipes) and from a shallow lake in Los Talas, Berisso (BA) (Scirpus californicus) [14]; in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: in vivo, formol-fixed.

Family LAGENOPHRYIDAE Bütschli, 1889

Lagenophrys Stein, 1852

Lagenophrys discoidea Kellicott, 1887

Recorded as epibiont on *Cyprinotus similis* (Ostracoda) from San Miguel del Monte Lake (BA) [20]. Observation techniques: *in vivo*.

Family OPERCULARIIDAE Fauré-Fremiet in Corliss, 1979

Opercularia Goldfuss, 1820

Opercularia cylindrata Wrzesniowski, 1870

Recorded in Capital Federal, epibiont on Cyclops sp. (Copepoda) [5] (checklist). Observation techniques: not indicated.

O. elongata Kellikott, 1884

Recorded in the periphyton (Myriophyllum quitense, Enteromorpha sp.) from San Miguel del Monte Lake (BA) [23]. Observation techniques: in vivo.

O. nutans (Ehrenberg, 1831) Stein, 1854

Recorded in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in the periphyton (Scirpus californicus, Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [17, 22, 46] (checklist). Cited as O. allensi by Modenutti & Claps [17]. Observation techniques: in vivo, Bouin-fixed, osmic acid vapours, formol-fixed.

Family OPHRYDIIDAE Ehrenberg, 1838

Ophrydium Bory de St. Vincent, 1824

Ophrydium naumanni Pejler, 1962

Recorded in the plankton from Moreno Oeste Lake (RN) [19, 36, 39, 41, 42, 44, 45, 47]. Observation techniques: in vivo, formol-fixed, Lugol-fixed.

O. versatile (O. F. Müller, 1786) Ehrenberg, 1830

Recorded in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton from lakes Lacar (NE), Gutiérrez (RN), Mascardi (RN), Guillelmo (RN) [66] (checklist). Observation techniques: fixed material.

Family VAGINICOLIDAE de Fromentel, 1874

Cothurnia Ehrenberg, 1831

Cothurnia annulata Stokes, 1885

Recorded in the periphyton (Myriophyllum quitense, Oedogonium sp.), on rotiferan lorigas, stalk of Campanella umbellaria, and in the benthos from San Miguel del Monte Lake (BA) [23, 46] (checklist). Observation techniques: in vivo, formol-fixed.

Cothurniopsis Stokes, 1893

Cothurniopsis valvata Stokes, 1893

Recorded in the plankton (on debris) and periphyton (Oedogonium sp.) from San Miguel del Monte Lake (BA) [23]. Observation techniques: in vivo.

Platycola Kent, 1882

Platycola decumbens Ehrenberg, 1830

Recorded in the periphyton from ponds in Los Talas, Berisso (BA), and Yalca and Chascomús lakes (BA), cited as P. striata and P. truncata [13]. Observation techniques: in vivo, mounted in glycerine.

P. dilatata Fromentel, 1874

Recorded in the periphyton from ponds in Los Talas, Berisso (BA), and Yalca and Chascomús lakes (BA), cited as P. tincta [13]. Observation techniques: in vivo, mounted in glycerine.

Pyxicola Kent, 1882

Pyxicola affinis Kent, 1882

Recorded in the periphyton from ponds in Los Talas, Berisso (BA), and Yalca and Chascomús lakes (BA) [13]. Observation techniques: in vivo, mounted in glycerine.

P. carteri Kent, 1882

Recorded in the periphyton from ponds in Los Talas, Berisso (BA), and Yalca and Chascomús lakes (BA), cited as Pyxicola cf. constricta [13]; in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); in the periphyton (Oedogonium sp.) from San Miguel del Monte Lake (BA), cited as P. limbata [23]. Observation techniques: in vivo, mounted in glycerine, formol-fixed.

Thuricola Kent, 1881

Thuricola folliculata Kent, 1881

Recorded in Capital Federal, in the plankton and periphyton (Vaucheria sp.) from ponds, streams, and rivers (not specifically indicated), cited as Cothurnia crystallina [5] (checklist). Observation techniques: not indicated.

T. innixa Stokes, 1882

Recorded in the periphyton (Myriophyllum quitense, Oedogonium sp.) from San Miguel del Monte Lake (BA) [23]. Observation techniques: in vivo.

T. kellicottiana (Stokes, 1887) Kahl, 1935

Recorded in the periphyton from ponds in Los Talas, Berisso (BA), and Yalca and Chascomús lakes (BA), cited as T. amphora [13]; in the periphyton (Myriophyllum quitense, Enteromorpha sp., Oedogonium sp.) and benthos from San Miguel del Monte Lake (BA) [23, 46]. Observation techniques: in vivo, mounted in glycerine, formol-fixed.

Vaginicola Lamarck, 1816

Vaginicola attenuata Fromentel, 1874

Recorded in the periphyton (Myriophyllum quitense, Oedogonium sp., Oscillatoria sp., diatoms) from San Miguel del Monte Lake (BA) [23]. Observation techniques: in vivo.

V. crystallina Ehrenberg, 1830

Recorded in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [22, 46] (checklist). Observation techniques: in vivo, formol-fixed.

V. ingenita (O. F. Müller, 1786) Kent, 1881

Recorded in San Miguel del Monte Lake (BA), in the periphyton (Myriophyllum quitense) and benthos [22, 46] (checklist). Observation techniques: in vivo, formol-fixed.

V. lagena Kahl, 1935

Recorded in the periphyton from ponds in Los Talas, Berisso (BA), and Yalca and Chascomús lakes (BA) [13]. Observation techniques: in vivo, mounted in glycerine.

V. tincta Ehrenberg, 1830

Recorded in the periphyton from ponds in Los Talas, Berisso (BA), and Yalca and Chascomús lakes (BA) [13]. Observation techniques: *in vivo*, mounted in glycerine.

Family VORTICELLIDAE Ehrenberg, 1838

Carchesium Ehrenberg, 1831

Carchesium polypinum (Linnaeus, 1758) Ehrenberg, 1830

Recorded in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton, periphyton, and mud samples from Río de la Plata [7] (checklist); Punta Atalaya, Magdalena (BA), in the periphyton (*Scirpus californicus*) from Río de la Plata [60]; in the periphyton from San Miguel del Monte Lake (BA) (*Ceratophyllum demersum*, *Scirpus californicus*) and from Río de la Plata (*Eichhornia crassipes*) in Punta Lara (BA) [17]; in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the plankton from Luján, Areco, and Ramallo rivers, and Cañada Honda (BA) [58] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours, formol-fixed.

C. spectabile Ehrenberg-Claparède & Lachmann, 1858

Recorded in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated), cited as *Carchesium lachmanni* [5] (checklist). Observation techniques: not indicated.

Intranstylum Fauré-Fremiet, 1904

Intrastylum invaginatum Stokes, 1886

Recorded near Poblet (BA), epibiont on Ostracoda from a temporary pond [29]. Observation techniques: in vivo, methyl-green-pyronin.

I. steini Wrzesniowsky, 1877

Recorded in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated), cited as *Epistylis steini* [5] (checklist). Observation techniques: not indicated.

Pseudovorticella Foissner & Schiffmann, 1975

Pseudovorticella difficilis (Kahl, 1933) Jankowski, 1976

Recorded in the periphyton (*Myriophyllum quitense*) from San Miguel del Monte Lake (BA) [22]. Observation techniques: *in vivo*.

P. monilata (Tatem, 1870) Foissner & Schiffmann, 1974

Recorded in the periphyton (*Ceratophyllum demersum*) from an artificial pool in La Plata (BA) [14]; in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: *in vivo*, formol-fixed.

P. nebulifera (O. F. Müller, 1786) Jankowski, 1976

Recorded in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not

specifically indicated) [5] (checklist); in the plankton and benthos from Río Primero (CO) [6]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist). Cited as *Vorticella nebulifera*. Observation techniques: *in vivo*, Bouin-fixed, osmic acid vapours.

Vorticella Linnaeus, 1767

Vorticella aquadulcis Stokes, 1887

Recorded near Poblet (BA), epibiont on subitaneous eggs of the colonial rotifer *Sinantherina* semibullata and in rewetted soil samples from a temporary pond [29]. Observation techniques: in vivo, methyl-green-pyronin.

V. campanula Ehrenberg, 1831

Recorded in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in Berisso (BA), in the periphyton (*Lemna* sp.) from a Palo Blanco pond and in Chascomús (BA), in Vitel stream [10]; epibiont on *Pomacea canaliculata*, *P. insularum*, *P. scalaris* (Gastropoda) from Middle Paraná River (SF) [72]; in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the plankton from Luján, Areco, and Baradero rivers, and Giles and de la Cruz streams (BA) [58] (checklist); in the periphyton (*Azolla* sp.) from Napostá Grande stream (BA) [18] (checklist); in the plankton from Río de la Plata estuary [37] (checklist); in the plankton from San Miguel del Monte Lake (BA) [54] (checklist); in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, methyl-green, neutral red, vital staining, Chatton-Lwoff silver staining [52], ferric hematoxylin, formol-fixed, silver impregnation after Fernández-Galiano [59], osmic acid vapours.

V. convallaria (Linnaeus, 1758) Linnaeus, 1767

Recorded in Capital Federal, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in Berisso (BA), in the periphyton from a Palo Blanco pond [10]; in Punta Atalaya, Magdalena (BA), in the periphyton (*Scirpus californicus*) from Río de la Plata [60] (checklist); in the periphyton from San Miguel del Monte (*Scirpus californicus*) and Chascomús (*Potamogeton striatus*) lakes, and from Río de la Plata (*Pistia stratiotes*) (BA) [14]; in La Plata (BA), in the plankton from Rodríguez stream [61] (checklist); in the plankton from Luján River (BA) [58] (checklist); in the plankton from Río de la Plata estuary [37] (checklist); in the periphyton (*Myriophyllum quitense*) and benthos from San Miguel del Monte Lake (BA) [46] (checklist); near Poblet (BA), in the plankton and periphyton (*Alternanthera philoxeroides*, *Ludwigia peploides*) from a temporary pond [29]; in the plankton from Salado River basin (BA) [49] (checklist). Observation techniques: *in vivo*, methyl-green, methyl-green-pyronin, neutral red, Chatton-Lwoff silver staining [52], ferric hematoxylin, silver staining [59], formol-fixed, osmic acid vapours.

V. halophila Stiller, 1941

Recorded near Poblet (BA), in the plankton and rewetted soil samples from a temporary pond [29]. Observation techniques: *in vivo*, methyl-green-pyronin.

V. longifilum Kent, 1881

Recorded near Poblet (BA), in the periphyton (Alternanthera philoxeroides) from a temporary pond [29] (checklist). Observation techniques: in vivo, methyl-green-pyronin.

V. microstoma Ehrenberg, 1830

Recorded in Capital Federal and Córdoba province, in the plankton and periphyton from ponds, streams, and rivers (not specifically indicated) [5] (checklist); in the plankton and benthos from Río Primero (CO) [6]; in the plankton, periphyton, and mud samples (not specifically indicated) from Río de la Plata [7] (checklist); in the periphyton (Azolla sp.) from Napostá Grande stream (BA) [18] (checklist); in the plankton from Río de la Plata estuary [37] (checklist); in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: in vivo, Bouin-fixed, osmic acid vapours, formol-fixed, methyl-green, neutral red, vital staining, silver staining [59].

V. natans Faure-Fremiet, 1924

Recorded in the plankton from Río de la Plata estuary [37] (checklist). Observation techniques: in vivo, methyl-green, neutral red, silver staining [59].

V. picta (Ehrenberg, 1831) Ehrenberg, 1838

Recorded in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [23,46] (checklist). Observation techniques: in vivo, formol-fixed.

V. pulchella Sommer, 1951

Recorded in the plankton (on debries) and periphyton (Myriophyllum quitense) from San Miguel del Monte Lake (BA) [23]; near Poblet (BA), in the periphyton (Ludwigia peploides) from a temporary pond [29]. Observation techniques: in vivo, methyl-green-pyronin.

V. rotunda Nenninger, 1948

Recorded in the periphyton (Myriophyllum quitense) from San Miguel del Monte Lake (BA) [23]. Observation techniques: in vivo.

V. striata Dujardin, 1841

Recorded in the plankton from Luján River (BA) [58] (checklist); in the periphyton (Myriophyllum quitense) and benthos from San Miguel del Monte Lake (BA) [46] (checklist). Observation techniques: in vivo, formol-fixed.

V. striata var. octava Stokes, 1885

Recorded in the periphyton from Las Viboras stream (Azolla filiculoides, Lemna sp.) in Magdalena (BA), and from Chascomús Lake (Ricciocarpus natans) (BA) [14]. Observation techniques: in vivo.

V. vestita Stokes, 1883

Recorded in the plankton from Río de la Plata estuary [37] (checklist). Observation techniques: in vivo, methyl-green, neutral red, silver staining [59].

Family ZOOTHAMNIIDAE Sommer, 1951

Zoothamnium Bory de St. Vincent, 1824

Zoothamnium ramosissimum Sommer, 1951

Recorded in the periphyton (Myriophyllum quitense) from San Miguel del Monte Lake (BA) [23]. Observation techniques: in vivo.

DISCUSSION

The total number of freshwater ciliates from Argentina comprises 209 species. Sampling efforts were mainly concentrated in the Buenos Aires province, although several surveys were conducted in Córdoba province as well. The Peritrichida are by far the most studied group of ciliates followed by Heterotrichida, Haptorida, and Sporadotrichida. The results in the present checklist show that a wide extension of Argentina is practically unexplored, and further investigations are needed to estimate the still undiscovered diversity of ciliates for the country.

Although most of the cited ciliates have cosmopolitan distributions, there are some examples of endemisms or species with restricted distributions in the South American ciliate biota. The most typical examples are 'flagships' or species that have a very distinctive morphology, size, and/or color to be overlooked if they were widely distributed [74]. One of such species is *Neobursaridium* gigas, which was discovered by Balech in vegetated ponds from Capital Federal and in Santa Fe province in Argentina [8]. Dragesco and Tuffrau [75] found this species later in subtropical Africa, which is of biogeographic interest since America and Africa were set appart almost 80 million years ago [76]. Another South American flagship species is Stentor araucanus, a blue-green planktonic ciliate described by Foissner and Wölfl in Chilean and Argentinean Andean lakes [55].

A great debate on the biogeography of protists is taking place nowadays and the ideas fall into two assumptions. Briefly, the 'ubiquity model' [77-79] assumes that all protists are distributed globally and have low degree of endemisms and low species numbers. This model is based on high gene flow between populations, high dispersion possibilities due to large population sizes, and hence, low rate of allopatric speciation. The 'moderate endemism model' [74, 80, 81] estimates a greater degree of endemism, with higher local radiation rates, and a notoriously undiscovered diversity. The continuing discovery of flagship species in different intensively sampled geographic regions is evidence in support for this model. Both hypotheses agree in the fact that most protists are cosmopolites, but fundamental problems in estimating diversity are undersampling and usually the study of very small samples, which leads to overlook the rare or encysted species [81]. On the other hand, the modern methodology employed for the accurate study of ciliates is rather difficult and also the use of old literature for species determination could lead to misidentifications. Considering that South America is almost unexplored for protist and most of the studies were carried out by means of live observations, a great diversity of ciliates is still awaiting to be discovered in this part of the world.

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REFERENCES

- [1] Lynn DH, Corliss JO. Ciliophora. In: *Microscopic anatomy of invertebrates. Vol. 1: Protozoa*, Harrison FW, Corliss JO (eds.), New York, Wiley-Liss Inc., 1991, 333-467.
- [2] Foissner W, Berger H, Schaumburg J. Identification and ecology of limnetic plankton ciliates. Informations berichte des Bayer, Landesamtes f
 ür Wasserwirtschaft, 3/99, 1999.
- [3] Foissner W. Biogeography and dispersal of micro-organisms: A review emphasizing protists. Acta Protozool 2006, 45:111-136.
- [4] de la Rua JM. Contribución al estudio de la microfauna de la Argentina. Protozoos. PhD Thesis, Universidad Nacional de Buenos Aires, 1911.
- [5] Seckt H. Estudios hidrobiológicos en la Argentina. Rev Univ Nac Córdoba 1924, 11:55-110.
- [6] Martínez-Bustos V. Contribución al conocimiento de la microbiología del agua del Río Tercero. Rev Univ Nac Córdoba 1933, 20:107-201.
- [7] Carbonell JJ. Some micrographic observations of the waters of the River Plate. Verh Internat Verein Limnol 1935, 7:513-516.
- [8] Balech E. Neobursaridium gigas n. gen. n. sp. de ciliado heterotrico. Physis B 1941, 21:29-34.
- [9] Ringuelet R. Sobre Folliculínidos de agua dulce de la Argentina. Physis B 1960, 21:315.
- [10] Cela AM. Algunos ciliados vinculados a la vegetación flotante. Physis B 1972, 31:559-577.
- [11] Vucetich MC. Presencia en la Argentina de Ascobius lentus Henneguy (Ciliata, Folliculinidae). Neotrópica 1972, 18:91-93.
- [12] Souto S. Contribución al conocimiento de los tintínnidos de agua dulce de la República Argentina. I. Río de la Plata y delta del Paraná. *Physis B* 1973, 32:249-254.
- [13] Vucetich MC, Escalante AH. Peritricos loricados de ambientes lénticos del área platense (Ciliata, Peritrichida). Neotrópica 1979, 25:187-194.
- [14] Claps MC, Modenutti BE. Contribución al conocimiento de los ciliados (Ciliophora Peritricha) dulceacuícolas de Argentina. II. *Limnobios* 1984, 2:581-585.
- [15] Claps MC, Modenutti BE. Ciliados dulceacuícolas de Argentina. IV. Suctorios del río Luján. Iheringia, Sér Zool 1988, 67:127-136.
- [16] Modenutti BE, Claps MC. Contribución al conocimiento de los ciliados dulceacuícolas de Argentina. I (Ciliophora-Suctoria). Neotrópica 1984, 30:121-124.
- [17] Modenutti BE, Claps MC. Ciliados dulceacuícolas de la Argentina, III: Ciliophora-Peritricha. Rev Asoc Cienc Nat Litoral 1986, 17:71-78.
- [18] Pettigrosso RE, Cazzaniga NJ. Registro de tres especies de Aspidisca (Ciliophora: Hypotrichida) en la Argentina. An Mus Hist Nat Valparaiso 1987, 18:5-12.
- [19] Modenutti BE. Presencia de Ophrydium naumanni Pejler (Ciliophora, Peritrichida) en lagos andinos rionegrinos. Neotrópica 1988, 36:99-103.
- [20] Claps MC, Sampóns, MR. First record of Lagenophrys discoidea Kellicott (Ciliophora, Peritricha, Lagenophryidae), ectocommensal of argentinian ostracods. Iheringia, Sér Zool 1994, 76:167-170.
- [21] Foggetta M, Boltovskoy A. Ciiiated Protozoa from oxygen depleted waters from Cassaffousth reservoir (Córdoba, Argentina). Rev Asoc Cienc Nat Litoral 1995, 26:25-31.
- [22] Zaleski M, Claps MC. First records of epiphytic limnetic ciliates from Argentina. Natura Neotropicalis 1999, 30:77-84.
- [23] Zaleski M, Claps MC. First record of some peritrichs ciliates for San Miguel del Monte pond (Buenos Aires, Argentina). *Gayana* 2001, 65:39-49.

- [24] Modenutti BE, Pérez GL. Planktonic ciliates from an oligotrophic south Andean lake, Morenito Lake (Patagonia, Argentina). Braz J Biol 2001, 61:389-395.
- [25] Dioni W. Un nuevo género de Folliculinidae de agua dulce: Botticula ringueleti nov. gen. nov. sp. del río Paraná Medio. Acta Zool Lilloana 1972, 29:304-313.
- [26] Souto S. Tintínnidos del Río de la Plata y su zona de influencia (Protozoa, Ciliata). Physis B 1974, 33:201-205.
- [27] Pettigrosso RE. Estudio taxonómico y ecológico de ciliados planctónicos (Ciliophora: Choreotrichida y Strombidiida) del estuario de Bahía Blanca, Argentina. PhD Thesis, Universidad Nacional del Sur, 2001.
- [28] Barría de Cao MS. Citología, bioecología y taxonomía de Tintinnida (Ciliophora). PhD Thesis, Universidad Nacional del Sur, 2002.
- [29] Küppers GC. Ciliados (Protozoa, Ciliophora) de un ambiente temporario subtropical pampásico: taxonomía y ecología. PhD Thesis, Universidad Nacional de La Plata, 2007.
- [30] Küppers GC, Lopretto EC, Claps MC. Pelagostrobilidium wilberti n. sp. (Oligotrichea, Choreotrichida): Morphology and morphogenesis. J Eukaryot Microbiol 2006, 53:477-484.
- [31] Küppers GC, Lopretto EC, Claps MC. Morphological aspects and seasonal changes of some planktonic ciliates (Protozoa) from a temporary pond in Buenos Aires Province, Argentina. *Panam JAS* 2006, 1:74-90.
- [32] Küppers GC, Claps MC, Lopretto EC. Description of *Notohymena pampasica* n. sp. (Ciliophora, Stichotrichia). *Acta Protozool* 2007, 46:221-227.
- [33] Küppers GC, Lopretto EC, Claps MC. Description of *Deviata rositae* n. sp., a new ciliate species (Ciliophora, Stichotrichia) from Argentina. *J Eukaryot Microbiol* 2007, 54:443-447.
- [34] Küppers GC, Claps MC, Lopretto EC. Ciliates (Protozoa, Ciliophora) from the dried sediments of a temporary pond from Argentina. Rev Mex Biodiv 2009, 80: in press.
- [35] Foggetta M. Prorodon taeniatus (Ciliophora Gymnostomata): Distribución espacio-temporal en el embalse Río Tercero. Limnobios 1987, 2:689-694.
- [36] Modenutti BE. Distribución de los ciliados planctónicos Ophrydium naumanni y Stentor araucanus en lagos oligotróficos andinos. Rev Soc Mex Hist Nat 1997, 47:79-83.
- [37] Rives CV. Protozoos como indicadores de contaminación. In: Calidad de las aguas de la franja costera del sur del Río de la Plata (San Fernando – Magdalena), AA-AGOSBA-ILPLA-SHN, Buenos Aires, 1997, 113-129.
- [38] Diéguez MC, Balseiro EG. Predation of Paradileptus elephantinus on rotifers. Verh Int Verein Limnol 1998, 27:2992-2995.
- [39] Queimaliños CP, Modenutti BE, Balseiro EG. Symbiotic association of the ciliate *Ophrydium naumanni* with *Chlorella* causing a deep chlorophyll-a maximum in an oligotrophic South Andes lake. *J Plankton Res* 1999, 21:167-178.
- [40] Modenutti BE, Balseiro EG, Moeller R. Vertical distribution and resistance to ultraviolet radiation of a planktonic ciliate Stentor araucanus. Verh Internat Verein Limnol 1998, 26:1636-1640.
- [41] Modenutti BE, Balseiro EG, Queimaliños CP. Ciliate community structure in two South Andean lakes: The effect of lake water on *Ophrydium naumanni* distribution. *Aquat Microb Ecol* 2000, 21:299-307.
- [42] Modenutti BE, Balseiro EG, Callieri C, et al. Increase in photosynthetic efficiency as a strategy of planktonic organisms exploiting deep lake layers. Freshwater Biol 2004, 49:160-169.

- [43] Modenutti BE, Balseiro EG, Callieri C, et al. Effects of UV-B and different PAR intensities on the primary production of the mixotrophic planktonic ciliate Stentor araucanus. Limnol Oceanogr 2005, 50:864-871.
- [44] Modenutti BE, Balseiro EG, Callieri C, Bertoni R. Light versus food supply as factors modulating niche partitioning in two pelagic mixotrophic ciliates. *Limnol Oceanogr* 2008, 53:446–455.
- [45] Balseiro EG, Modenutti BE, Queimaliños CP. Feeding of *Boeckella gracilipes* (Copepoda, Calanoida) on ciliates and phytoflagellates in an ultraoligotrophic Andean lake. *J Plankton Res* 2001, 23:849-857.
- [46] Zaleski M, Claps MC. Ciliados indicadores de la calidad de agua de la laguna San Miguel del Monte (Buenos Aires). Diversidad y Ambiente 2000, 1:45-51.
- [47] Modenutti BE, Balseiro EG. Mixotrophic ciliates in an Andean lake: Dependence on light and prey of an Ophrydium naumanni population. Freshwater Biol 2002, 47:121-128.
- [48] Kogan M. Estudio de la composición específica, abundancia y distribución especial del microzooplancton (protozoos y micrometazoos) en el estuario del Río de la Plata (Argentina-Uruguay). PhD Thesis, Universidad de Buenos Aires, 2005.
- [49] Claps MC, Gabellone NA, Neschuk NC. Influence of regional factors on zooplankton structure in a saline lowland river: The Salado River (Buenos Aires, Argentina). River Res Appl 2009, 25:453-471.
- [50] Lynn DH. The ciliated protozoa. Characterization, classification, and guide to the literature. 3rd Edn, Canada, Springer, 2008.
- [51] Wilbert N. Eine verbesserte Technik der Protargolimprägnation für Ciliaten. Mikrokosmos 1975, 64:171-179.
- [52] Corliss JO. Silver impregnation of ciliated protozoa by the Chatton-Lwoff technique. Stain Tech 1953, 28:97-100.
- [53] Carey P. Marine interstitial ciliates. An illustrated key. London, Chapman & Hall, 1992.
- [54] Benítez HH. Estructura y dinámica del zooplancton de una laguna pampásica moderadamente eutrófica (San Miguel del Monte): factores de control. PhD Thesis, Universidad Nacional de La Plata, 2008.
- [55] Foissner W, Wölfl S. Revision of the genus Stentor Oken (Protozoa, Ciliophora) and description of S. araucanus nov. spec. from South American lakes. J Plankton Res 1994, 16:255-289.
- [56] Foissner W. Basic light and scanning electron microscopic method for taxonomic studies of ciliated protozoa. Europ J Protistol 1991, 27:313-330.
- [57] Gaggero P. Presencia de Stentor coeruleus Ehrbg. en La Plata. Memorias Jardín Zoológico La Plata 1928, 3:25-28.
- [58] Modenutti BE. Zooplancton de ambientes lóticos de la subcuenca delta del río Paraná, Buenos Aires, Argentina. *Iheringia*, Sér Zool 1991, 71:67-80.
- [59] Fernández-Galiano D. Silver impregnation of ciliated protozoa: Procedure yielding good results with the pyridinated silver carbonate method. Trans Am Microsc Soc 1976, 94:557-560.
- [60] Claps MC. Zooperifiton en Scirpus (Scirpus) californicus (Meyer) Steud. (Río de la Plata Punta Atalaya). Neotrópica 1984, 30:79-88.
- [61] Modenutti BE. Variaciones en la diversidad del zooplancton del arroyo Rodríguez (prov. de Buenos Aires). Rev Asoc Cienc Nat Litoral 1987, 18:61-70.
- [62] Song W, Warren A, Hu X. Free-living ciliates in the Bohai and Yellow seas, China. Beijing, Science Press, 2009.

- [63] Song W, Warren, Wang M. Redescriptions of three marine ciliates, Strombidium elegans Florentin, 1901, Strombidium sulcatum Claparede & Lachmann, 1859 and Heterostrombidium paracalkinsi Lei, Xu & Song, 1999 (Ciliophora, Oligotrichida). Europ J Protistol 2000, 36:327-342.
- [64] Foissner W, Berger H, Blatterer H, Kohmann F. Taxonomische und ökologische Revision der Ciliaten des Saprobiensystems - Band IV: Gymnostomates, Loxodes, Suctoria. Informationsberichte des Bayer, Landesamtes für Wasserwirtschaft 1/95, 1995.
- [65] Frenzel J. Über einige merkwurdige Protozoen Argentiniens. Z wiss Zool 1891, 53:334-360.
- [66] Thomasson K. Araucanian Lakes: Plankton studies in North Patagonia with notes on the terrestrial vegetation. Acta Phytogeogr Suecica 1963, 47:1-139.
- [67] Ardohain DM. Respuesta del zooplancton en su estructura y dinámica a factores clave en una laguna arreica (provincia de Buenos Aires). PhD Thesis, Universidad Nacional de La Plata, 2008.
- [68] Dragesco J, Dragesco-Kernéis A. Ciliés libres de l'Afrique intertropicale. Introduction á la connaissance et á l'étude des ciliés. Faune Tropicale 1986, 26:1-159.
- [69] Foissner W, Berger H, Kohmann F. Taxonomische und ökologische Revision der Ciliaten des Saprobiensystems - Band III: Hymenostomata, Prostomatida, Nassulida. Informationsberichte des Bayer, Landesamtes für Wasserwirtschaft 1/94, 1994.
- [70] Bauer DE, Gómez N, Hualde PR. Biofilms coating Scirpus californicus as indicators of water quality in the Río de la Plata estuary (Argentina). Environ Monit Assess 2007, 133:309-320.
- [71] Juárez J, Villagra de Gamundi A, López Z, et al. Consideraciones sobre la taxocenosis de ciliados en un proceso de tratamiento de un efluente citrícola (Tucumán – Argentina). Ecología en Bolivia 2002, 37:59-69.
- [72] Di Persia DH, Radici de Cura MS. Algunas consideraciones acerca de los organismos epibiontes desarrollados sobre Ampullariidae. *Physis B* 1973, 32:309-319.
- [73] Thomasson K. Studies on South American freshwater plankton from Tierra del Fuego and Valdivia. Acta Horti Gotob 1955, 19:193-225.
- [74] Foissner W. Biogeography and dispersal of micro-organisms: A review emphasizing protists. *Acta Protozool* 2006, 45:111-136.
- [75] Dragesco J, Tuffrau M. Neobursaridium gigas Balech, 1941, Cilié Holotriche Hymenostome pantropical. Protistologica 1967, 3:133-146.
- [76] Dragesco J. 50 years of study in Protozoa. Personal experiences. Trav Mus Natl Hist Nat Grigore Antipa 1998, 40:7-25.
- [77] Finlay BJ, Corliss JO, Esteban G, Fenchel T. Biodiversity at the microbial level: The number of free-living ciliates in the biosphere. Rev Biol 1996, 71:221-237.
- [78] Finlay BJ, Esteban GF, Fenchel T. Protozoan diversity: Converging estimates of the global number of free-living ciliate species. *Protist* 1998, 149:29-37.
- [79] Finlay BJ, Esteban GF. Ubiquitous dispersal of free-living microorganisms. In: Microbial diversity and bioprospecting, Bull AT (ed.). Washington, DC, ASM Press, 2004, 216-224.
- [80] Foissner W. Protist diversity: Estimates of the near-imponderable. Protist 1999, 150:363-368.
- [81] Foissner W. Protist diversity and distribution: Some basic considerations. *Biodivers Conserv* 2008, 17:235-242.